

**ISSN 0189-6016©2009**MICROBIAL BURDEN OF SOME HERBAL ANTIMALARIALS MARKETED AT ELELE,  
RIVERS STATE**\*<sup>1</sup>Tatfeng Y.M, <sup>2</sup>Olama E.H and <sup>1</sup>Ojo T.O**<sup>1</sup>Department of Medical Laboratory Sciences, Niger Delta University, Wilberforce Island,  
Bayelsa state.<sup>2</sup>Department of Medical Laboratory Sciences, Madonna University, Elele, Rivers state**\*Email:** [youchou@yahoo.com](mailto:youchou@yahoo.com)**Abstract**

Herbal antimalarials still remain an alternative to our traditional communities who can not afford orthodox antimalarials. This study was aimed at investigating the microbial quality of six herbal antimalarials using standard microbiological methods. Of the six preparations analyzed, “schnapps”, palm wine and water were the media of preparation; the water base preparations recorded higher microbial load. The mean microbial load was  $159.5 \times 10^5$  cfu/ml and  $217.4 \times 10^2$  cfu/ml in water and alcohol base preparations respectively. The microbial profile of the preparations showed that the schnapps base preparations were predominantly contaminated with *Bacillus* sp (Aerobic spore bearers) and *Mucor* spp. The palm wine preparation harboured *Bacillus* sp, yeasts and *Mucor* spp while the water base preparations had several isolates such as *Staphylococcus epidermidis*, *Pseudomonas aeruginosa*, *Escherichia coli* O157H7, *Proteus mirabilis*, *Enterococcus faecalis*, *Serratia marcescens*, *Staph. aureus*, *Bacillus* spp and *Mucor* spp. Conclusively, this study underlines the public health importance of these preparations given the high burden of such human pathogen as *Ecoli* O157H7, *Ps aeruginosa*, *Staph aureus*, etc. in the preparations.

**Key words:** antimalarials, herbal, microbial, Elele**Introduction**

Malaria is a major dreaded disease of humans causing hundreds of millions of morbidity with increasing rate of mortality. The number of people dying from malaria is now higher than it was 30 years ago (Joy *et al.*, 2003). Traditional medicines have been used to treat malaria for thousands of years and are the source of the two main groups (artemisinin and quinine derivatives) of modern antimalarial drugs. With the problems of increasing levels of drug resistance and the inability of a large number of patients being able to afford and access effective antimalarial drugs, traditional medicines could be an important and sustainable source of treatment (Wilcox and Bodeker, 2004). Resistance has spread so fast that it now represents a serious threat to global public health (World Health Organization, 2003). Scientists now agree that the most effective treatment against malaria is a combination of drugs using artemisinin derivatives which are highly potent extracts of the Chinese plant *Artemisia annua*. Artemisinin based combination therapy (ACT) is the quickest and most reliable way of clearing malaria infection and has been shown to protect each individual drug from resistance (World Health Organization, 2002). Herbal therapy has been used in the treatments of many ailments, including malaria. Various plant parts singly or a combination of

a number of plants have been employed in the treatment of malaria; some plants which have been reported include *Azadirachta indica*, *Morinda lucida*, *Alstonia boonei*, *Enantia chlorantha*, *Crossopteryx febrifuga*, *Tithonia diversifolia* etc. (Tella, 1977; Ade 1983; Obih and Makinde, 1985; Makinde *et al.*, 1987; Elufioye and Agbedahunsi, 2004). In this work, we investigated the microbial quality of some antimalarials marketed at Elele, Rivers State.

## Materials and Methods

### Study design

Six preparations of antimalarials herbal preparations were obtained from different marketers at Elele, Rivers state, Nigeria. The products were collected in sterile containers to avoid contamination from external source for microbial analysis. The composition of the preparations was as given below;

**Preparation 1:** chopped Roots of *Azadirachta indica* A. Juss family Meliaceae soaked in Schnapps.

**Preparation 2:** Sliced leaves *Cymbopogon citratus* (DC.) Stapf., family Graminae soaked in alcohol Schnapps.

**Preparation 3:** Sliced stem barks of *Azadirachta indica* A. Juss family Meliaceae soaked in palm wine

**Preparation 4:** Sliced stem barks of *Azadirachta indica* A. Juss family Meliaceae soaked in water

**Preparation 5:** Leaves of *Ocimum gratissimum* Linn. (Labiatae), *Psidium guajava* Linn. (Myrtaceae), *Cymbopogon citratus* (DC.) Stapf., Graminae), *Azadirachta indica* A. Juss (Meliaceae), *Mangifera indica* Linn. (Anarcadiaceae) and unripe leaf of *Carica papaya* Linn. (Caricaceae) all cut into pieces, boiled in water, allowed to cool and sieved into a clean container.

**Preparation 6:** Leaves of *Cymbopogon citratus* (DC.) Stapf., Graminae), leaves of *Azadirachta indica* A. Juss (Meliaceae), leaves of *Mangifera indica* Linn. (Anarcadiaceae) and unripe bark of the fruit of *Carica papaya* Linn. (Caricaceae) and leaves of *Vernonia amygdalina* Del. (Compositae) boiled in a pot till it changed colour, allowed to cool and then sieved into a clean container.

### Sample Processing

#### Microbial Isolation and Identification

The isolation of microbes was done culturally using Eosin Methylene Blue agar, Potato Dextrose agar, Blood agar and Salmonella-Shigella agar plates and microbial identification was carried out using standard microbiological techniques, this includes Gram staining of the isolates, biochemical testing i.e. coagulase, catalase, citrate utilization, indole production, oxidase test, sugar fermentation tests, serology for *E. coli* O157H7 using and lacto phenol cotton blue preparations.

#### Total Viable Count

A ten fold dilution was carried out on the six samples. The dilution was made as follows, 9 ml of sterile water was transferred in each of 6 rows of 10 sterile test tubes, 1 ml of each of the samples was transferred in the first test tubes on each row containing sterile distilled water and was mixed properly. One (1) ml from the first tube on each row was transferred to the 2<sup>nd</sup> test tube and was mixed by shaking to obtain a 1/10 dilution. This exercise was carried though the tenth tube where 1 ml of the mixture was eventually discarded away to obtain dilutions of 1/10<sup>2</sup>, 1/10<sup>3</sup>, 1/10<sup>4</sup>...1/10<sup>10</sup>. One (1) ml from each test tube was transferred into a sterile petri dish and molten nutrient agar was added, mixed well and incubated aerobically at 37 °C. Count was made on the plate showing evenly distributed and discrete colonies.

## Results

Of the six preparations analyzed, schnapps, palm wine and water were the media of preparation, the 2 alcohol base preparations had count of  $0.2 \times 10^2$  cfu/ml and  $2.0 \times 10^2$  cfu/ml, the only palm wine preparation had a count of  $6.5 \times 10^4$  cfu/ml while the 3 water based preparations had count of  $5.6 \times 10^5$  cfu/ml,  $3.1 \times 10^5$  cfu/ml and  $4.7 \times 10^7$  cfu/ml each. The microbial profile showed that the schnapps base preparations were mostly contaminated with *Bacillus* spp (Aerobic spore bearers) and *Mucor* spp, the palm wine preparation harboured *Bacillus* spp, yeasts and *Mucor* spp while the water based preparations had several microbial isolates such as *Staph. epidermidis*, *Pseudomonas aeruginosa*, *E. coli* 0157, *Proteus mirabilis*, *Enterococcus faecalis*, *Serratia marcensces*, *Staph. aureus*, *Bacillus* spp and *Mucor* spp (Table 1).

**Table 1:** Total viable count and distribution of microbial isolates by sample.

| Samples                | 1                         | 2                         | 3                               | 4                             | 5                             | 6                             |
|------------------------|---------------------------|---------------------------|---------------------------------|-------------------------------|-------------------------------|-------------------------------|
| Microbial Load(cfu/ml) | $0.2 \times 10^2$         | $2.0 \times 10^2$         | $6.5 \times 10^4$               | $5.6 \times 10^5$             | $3.1 \times 10^5$             | $4.7 \times 10^7$             |
| Isolates               | <i>Bacillus</i> spp (ASB) | <i>Bacillus</i> spp (ASB) | <i>Bacillus</i> spp (ASB)       | <i>Staph. epidermidis</i>     | <i>Pseudomonas aeruginosa</i> | <i>Pseudomonas aeruginosa</i> |
|                        | <i>Mucor</i> spp          | <i>Mucor</i> spp          | <i>Mucor</i> spp<br>Yeast cells | <i>Bacillus</i> spp           | <i>Proteus mirabilis</i>      | <i>Staph. Aureus</i>          |
|                        |                           |                           |                                 | <i>Pseudomonas aeruginosa</i> | <i>Bacillus</i> (ASB)         | <i>E. coli</i>                |
|                        |                           |                           |                                 | <i>E. coli</i> 0157           | <i>Serratia marcensces</i>    | <i>Staph epidermidis</i>      |
|                        |                           |                           |                                 | <i>Proteus mirabilis</i>      | <i>Mucor</i> spp              | <i>Bacillus</i> spp           |
|                        |                           |                           |                                 | <i>Mucor</i> spp              |                               | <i>Mucor</i> spp              |
|                        |                           |                           |                                 | <i>Aspergillus niger</i>      |                               | <i>Enterococcus faecalis</i>  |

## Discussion

Herbal antimalarials are herbal drugs used in the form of concoction and decoction from roots, stems and flowers from various plants which is believed to cure malaria. They have some risks associated with their use. Findings from this study showed that the antimalarials studied were contaminated with large number of pathogenic organisms of public health importance. *Bacillus* spp and *S. aureus* had the highest frequency of occurrence. This was similar to previous findings reported by World Health Organization (WHO, 1998). The occurrence of isolates such as *E. coli* O157H7 and *Enterococcus faecalis* is of serious medical importance as its presence could indicate a potential fecal contamination. Both water and alcohol base preparations harboured organisms; however, "Schnapps" base preparations recorded lower microbial load than the water base preparations which recorded load above acceptable levels. Obviously, Schnapps contains some level of alcohol at concentration which could have some inhibitory activities on some microorganisms. Water base preparations contamination could result from the water used for the preparation, the traditional healers could have been the source of contamination as the hygienic requirement may not have been met. Soil could also be a source of contamination as medicinal plant materials normally carry a large number of microbes originating from the soil. The usage of alcohol containing medium in the preparations of these products remains worrisome in our communities where such are used also as prophylaxis by both children and adults bearing in mind the effect of alcohol on liver functions.

Clinical observations on traditional remedies are feasible and useful. Some herbal remedies may be safe and effective for the treatment of malaria. Nevertheless, better evidence from randomised clinical

trials is needed before herbal remedies can be recommended on a large scale. Preventing children's deaths is the key objective of any malaria control programme. Once a remedy has been shown to be safe and effective for uncomplicated malaria in adults, studies on mortality in children would be the necessary next step. It has already been shown that mortality can be reduced in the under 5s by training mothers to recognise malaria and to give early treatment (Kidane and Morrow, 2000).

In conclusion, some herbal antimalarials may be helpful in the management of malaria fever; serious attention should be given to this alternative branch in medicine so that its limitations do not overshadow its benefits.

## References

1. Ade, M.A. (1983). Antimalaria and Anti-lymphocytotoxic properties of *Azadirachta indica* (Dongoyaro). *Research into African Medicinal Plants Newsletter*. **27 (9 & 10)**: 85.
2. Elufioye, T.O and Agbedahunsi, J.M. (2004). Antimalaria activities of *Tithonia diversifolia* (Asteraceae) and *Crossopteryx febrifuga* (Rubiaceae) on mice *in vivo*. *J. Ethnopharmacol.* **31**:1 – 5.
3. Joy, D., Feng, X. and Mu, J. (2003). Early origin and recent expansion of *Plasmodium falciparum*. *Science*. **300(5617)**: 318 – 21.
4. Kidane, G. and Morrow, R.H. (2000). Teaching mothers to provide home treatment of malaria in Tigray, Ethiopia: a randomised trial. *Lancet*. **356**: 550-5.
5. Makinde, J.M., Obih, P.O., and Jimoh, A.A. (1987). Effects of *Solanum erianthum* Aqueous leaf extract on *Plasmodium berghei berghei* in mice. *Afr. J. of Med. Med. Sc.* **16**:193 – 196.
6. Obih, P.O. and Makinde, J.M (1985). Effect of *Azadirachta indica* on *Plasmodium berghei* in Mice. *Afr. J. of Med. Med. Sc.* **14**:51 – 54.
7. Tella, A. (1977).The effects of *Azadirachta indica* in Acute plasmodium berghei malaria. *Nig. Med. Journal.* **7(3)**: 258-259.
8. Wilcox, M.L. and Bodeker, G. (2004). Traditional herbal medicines for malaria. *BMJ.* **329**:1156-1159.
9. World Health Organization. (1998). Quality control methods for the medicinal plants: determination of microorganisms. *WHO Geneva technical report.* 64-73.
10. World Health Organization. (2002). Report of a WHO Technical Consultation, 4 – 5. Geneva.
11. World Health Organization. (2003). WHO Press Release WHO/85 11 November 2003.